Assay Training: Xpert® Xpress CoV-2/Flu/RSV *plus*

For Use with GeneXpert® Dx or GeneXpert Infinity Systems

CE IVD In Vitro Diagnostic Medical Device

302-7366 Rev. C November 2022

Training Agenda

- Xpert® Xpress CoV-2/Flu/RSV *plus*
  - Reagents
  - Specimen collection, storage, & handling
  - Kit storage and handling
  - Cartridge preparation
  - Quality controls
  - Results analysis
- Discussion

Training Objectives

At the end of the training, users will be able to:

- Properly store and handle the Xpert® Xpress CoV-2/Flu/RSV *plus* kit
- Follow proper laboratory safety precautions
- Collect and store appropriate specimen(s)
- Prepare a cartridge and run the Xpert Xpress CoV-2/Flu/RSV *plus* test
- Report the various software generated results
- Understand the Xpert Xpress CoV-2/Flu/RSV *plus* control strategy

The Cepheid Solution

- Detection of SARS-CoV-2, Flu A, Flu B, RSV RNA
- On-board internal controls for each sample
- Probe Check Control (PCC)
- Sample Processing Control (SPC)
- Closed cartridge system minimizes risk of contamination
- EAT (Early Assay Termination for SARS-CoV-2 ADF only)
- On-demand results
- Random access

Intended Use

- The Xpert® Xpress CoV-2/Flu/RSV *plus* test, performed on the GeneXpert® Instrument Systems, is a multiplexed real-time RT-PCR test intended for use in the simultaneous, *in vitro* qualitative detection and differentiation of RNA from SARS-CoV-2, influenza A, influenza B, and/or respiratory syncytial virus (RSV) in either nasopharyngeal swab or anterior nasal swab specimens collected from individuals with signs and/or symptoms of respiratory viral infection.
- SARS-CoV-2, influenza, A, influenza B and RSV RNA identified by this test are generally detectable in upper respiratory specimens during the acute phase of infection. Positive results are indicative of the presence of the identified virus, but do not rule out bacterial infection or co-infection with other pathogens not detected by the test.
- Clinical correlation with patient history and other diagnostic information is necessary to determine patient infection status. The agent detected may not be the definite cause of disease.
- Negative results do not preclude SARS-CoV-2, influenza A virus, influenza B virus and/or RSV infection and should not be used as the sole basis for treatment or other patient management decisions. Negative results must be combined with clinical observations, patient history, and/or epidemiological information.

Good Laboratory Practice Review

Personal Protective Equipment (PPE)	• Wear clean lab coats, safety glasses, and gloves • Change gloves between processing samples
Lab Bench Area	• Clean work surfaces routinely with: ✓ 1:10 dilution of household bleach* ✓ 70% Ethanol Solution • After cleaning, ensure work surfaces are dry
Specimens, Samples, and Kits Storage	• Store specimens and sample away from kit to prevent contamination
Equipment	• Use filtered pipette tips when recommended • Follow the manufacturer's requirements for calibration and maintenance of equipment

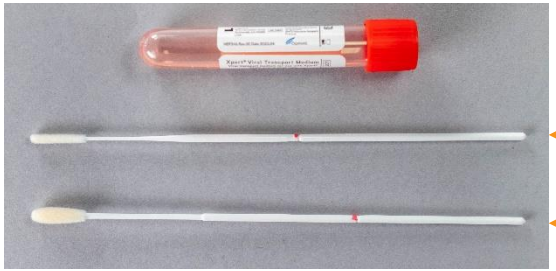
* Final active chlorine concentration should be 0.5% regardless of the household bleach concentration in your country.

Cepheid

Specimen Collection, Storage and Handling

Specimen Collection

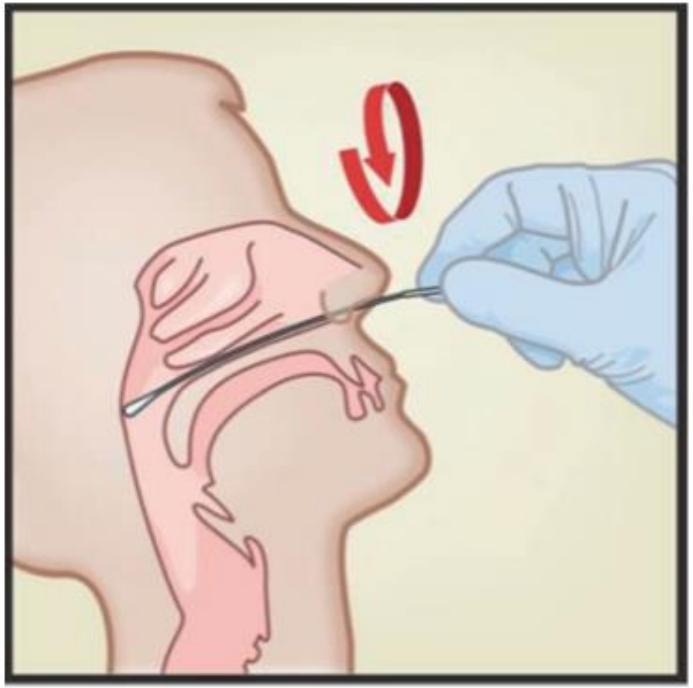
Specimen Type	Storage Type
Nasopharyngeal swab	Place specimen into 3 mL of viral transport medium, 3 mL of saline, or 2 mL of eNAT™
Anterior nasal swab	



Refer to the WHO Laboratory Biosafety Guidance Related to the Coronavirus Disease 2019 (COVID-19).
https://www.who.int/publications-detail/laboratory-biosafety-guidance-related-to-coronavirus-disease-2019-(covid-19)

Specimen Collection- Nasopharyngeal Swab

1. Insert the swab into either nostril, passing it into the posterior nasopharynx.
2. Rotate swab by firmly brushing against the nasopharynx several times.
3. Remove and place the swab into the transport tube.
4. Break swab at the indicated break line.
5. Cap the specimen collection tube tightly.



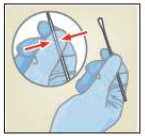
Specimen Collection- Nasopharyngeal Swab

Nasopharyngeal Specimen Collection

1. Open the package that contains the swab and transport medium tube. Set the tube aside before collecting the specimen.

2. Open the swab wrapper and remove the swab, taking care not to touch the tip of the swab to any surface.

3. Hold the swab in your hand, pinching in the middle of the swab shaft on the scoreline.



4. Gently insert the swab into the nostril until you touch the posterior nasopharynx.

   Rotate swab several times.

5. Remove the cap from the tube.
   Insert the swab into the transport medium.

6. Break the swab shaft against the side of the tube at the scoreline.

   Avoid splashing contents on the skin. Wash with soap and water if exposed.

7. Replace the cap on the tube and close tightly.

301-6052, Rev. H September 2021

Cepheid
A better way.

Cepheid

Specimen Collection- Nasal Swab

1. Insert the nasal swab 1 to 1.5 cm into the nostril.
2. Rotate the swab against the inside of the nostril for 3 seconds while applying pressure with a finger to the outside of the nostril.
3. Repeat on the other nostril with the same swab.
4. Remove and place the swab into the transport tube.
5. Break swab at the indicated break line.
6. Cap the specimen collection tube tightly.

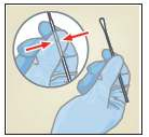
Specimen Collection- Nasal Swab

Nasal Swab Specimen Collection

1. Open the package that contains the swab and transport medium tube. Set the tube aside before collecting the specimen.

2. Open the swab wrapper and remove the swab, taking care not to touch the tip of the swab to any surface.

3. Hold the swab in your hand, pinching in the middle of the swab shaft on the scoreline.



4. Rotate swab against the inside of the nostril for 3 seconds while applying pressure with a finger to the outside of the nostril.

   Do not insert the swabs more than 1-1.5 cm.

5. Repeat Step 4 on the other nostril with the same swab.

   To avoid specimen contamination, do not touch the swab tip to anything after collecting the specimen.

6. Remove the cap from the tube.
   Insert the swab into the transport medium.

7. Break the swab shaft against the side of the tube at the scoreline.

   Avoid splashing contents on the skin. Wash with soap and water if exposed.

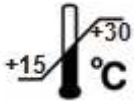
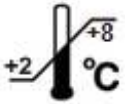
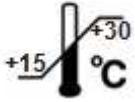
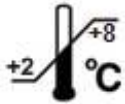
8. Replace the cap on the tube and close tightly.

301-9057, Rev. F September 2021

Cepheid
A better way.

Cepheid

Specimen Transport and Storage

Sample Type	Transport and Storage Conditions	
Transport tube containing nasopharyngeal swab or nasal swab viral transport medium or saline*		≤ 48 hours
		≤ 7 days
Transport tube containing nasopharyngeal swab or nasal swab eNAT™		≤ 48 hours
		≤ 6 days

*Samples collected into saline should not be frozen.

Kit Storage and Handling

Xpert® Xpress CoV-2/Flu/RSV *plus* Requirements

GeneXpert® Dx and GeneXpert Infinity System

- GeneXpert Dx software version 4.7b or higher
- For GeneXpert Infinity-80 and Infinity-48s systems: Xpertise software version 6.4b or higher

Test Kits

- XP3COV2/FLU/RSV-10

Materials Required but not Provided

- Nylon flocked swab (Copan P/N 502CS01, 503CS01) or equivalent
- Viral transport medium, 3 mL (Copan P/N 330C) or equivalent
- 0.85-0.9% (w/v) saline, 3 mL
- Nasopharyngeal and Nasal (Copan Part Number 50 units : 305C & 346C) or equivalent

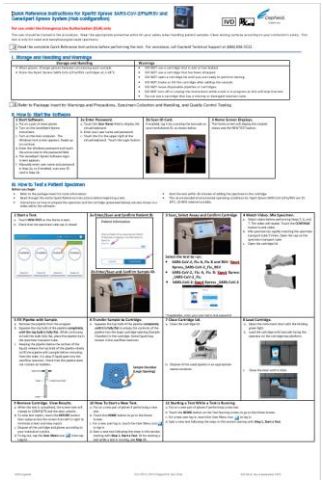
Optional

- Uninterruptible Power Supply/ Surge Protector
- Printer

Kit Components

Xpert® Xpress CoV-2/Flu/RSV *plus*

Catalog Number	**XP3COV2/FLU/RSV-10**
Tests per Kit	10
Transfer Pipettes	Instructions to locate (and import) the ADF and EUA documentation such as the Product Insert on www.cepheid.com
Storage Temperature	2-28°C



The kit also includes printed copies of the Quick Reference Instructions, which should **only** be used with the GeneXpert® Xpress System.

Cepheid.
For *in Vitro* Diagnostic use

IVD CE 302-7086, Rev. A September 2021

Cartridges contain chemically hazardous substances-please see Instructions for Use and Safety Data Sheet for more detailed information.

Cepheid

Kit Storage and Handling

- Store test kits at 2-28ºC. Do not use expired cartridges.
- Each single-use cartridge is used to process one test. Do not reuse processed cartridges.
- Do not open a cartridge until ready to use.
- Start the test within 30 minutes of adding the sample to the cartridge.
- To avoid cross contamination during sample handling steps, change gloves between samples.

Cartridge Preparation

Warning and Precautions

- Do not shake the cartridge.
- Do not use a cartridge…
  - if it appears wet, has leaked, or if the lid seal appears to have been broken.
  - if it appears damaged.
  - that has been dropped after removing it from packaging.
  - that has been dropped or shaken after you have added the sample.
  - that has a damaged reaction tube.
  - that has been used; each cartridge is single-use to process one test.
  - that has expired.
- Do not reuse pipettes.
- Do not reuse swabs.

Proper Cartridge Handling Techniques

- Do not touch the reaction tube.
- Keep the cartridge upright after seal has been broken.
- Do not tilt when scanning the cartridge.

Xpert® Xpress CoV-2/Flu/RSV *plus* Cartridge Preparation

Check if all items below are present:
1. Transport media containing swab (if applicable)
2. Patient name or identifier on the tube
3. Cartridges and transport media are within the expiration date

Good Laboratory Practices:
- Wear clean gloves, safety glasses, and lab coats.
- Change gloves between samples.
- Clean work surface with 1:10 dilution of bleach followed by 70% ethanol solution.

Xpert® Cartridge Preparation

- Xpert Xpress SARS-CoV-2
- Xpert Xpress SARS-CoV-2/Flu/RSV
- Xpert Xpress CoV-2/Flu/RSV *plus*

Refer to the package insert for detailed instructions, precautions, and warnings.

For a copy of the SDS, visit www.cepheid.com or www.cepheidinternational.com

Contact information for all Cepheid Technical Support offices is available on our website: www.cepheid.com/en/CustomerSupport

Cepheid. A better way.

1 Take one Xpert cartridge for each sample.

2 Rapidly invert the tube 5 times.

3 Open the cartridge lid.

4 Using a clean 300 µL pipette (supplied), transfer 300 µL (one draw), of the sample to the opening of the cartridge.

5 Close the cartridge lid.

6 Start the test within the timeframe specified in the package insert.

 CE IVD *In Vitro* Diagnostic Use. *In vitro* medical diagnostic device. May not be available in all countries.

302-3816, Rev. B September 2021

Cepheid.

Xpert® Xpress CoV-2/Flu/RSV *plus* Cartridge Preparation

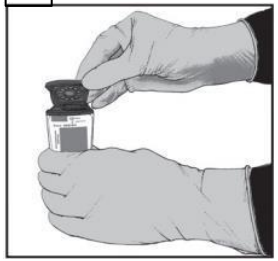
1

Take one Xpert cartridge for each sample.

2

Rapidly invert the tube 5 times.

3



Open the cartridge lid.

4

Using a clean 300 µL pipette (supplied), transfer 300 µL (one draw) of the sample to the cartridge.

5

Close the cartridge lid.

6

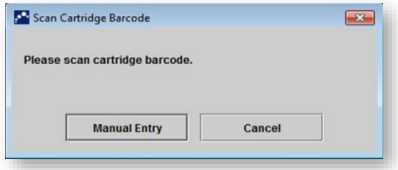
Start the test within the timeframe specified in the Instructions For Use.

Run a Test on GeneXpert® Dx

1 Create a test.

Start the test within 1 hour after adding the sample to the cartridge.

2 Scan barcode for Patient and/or Sample ID.



Do not click on Manual Entry or Cancel.

3 Scan the cartridge.

For complete details on how to run a test, refer to the Package Insert and the GeneXpert Dx or Xpertise Operator Manual.

Cepheid

Run a Test on GeneXpert® Dx (continued)

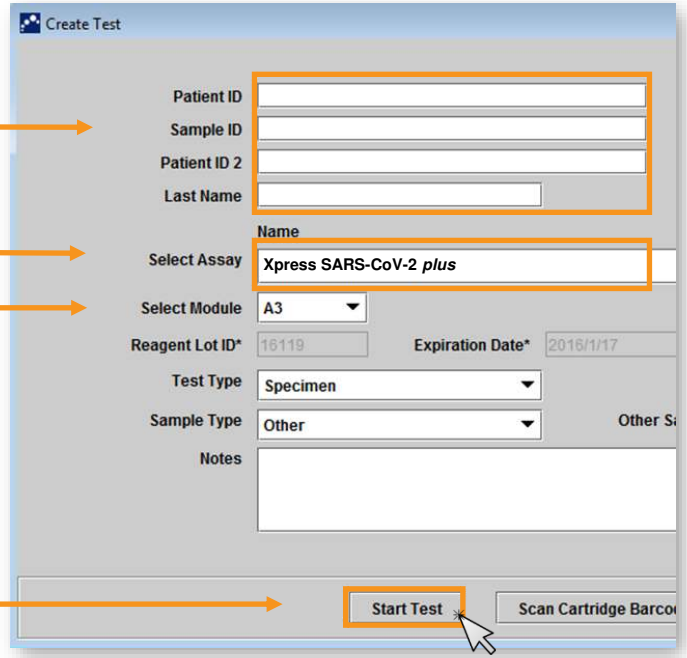
4 Complete the fields as required.

5 Select the assay protocol.

6 The module is selected automatically.

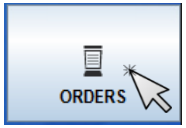
7 Click on Start Test.

8 A green light will flash on the module.
Load the cartridge into module and close the door.



Run a Test on GeneXpert® Infinity

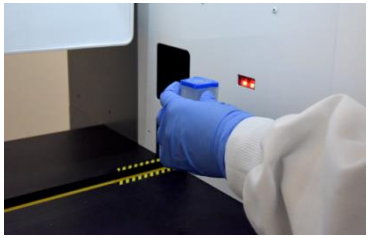
1 Create a test.



Place the cartridge on the conveyor within 15 minutes of adding the sample.

2 Scan barcode for Patient and/or Sample ID.

3 Scan the cartridge.



For complete details on how to run a test, refer to the Package Insert and the GeneXpert Dx or Xpertise Operator Manual.

Cepheid

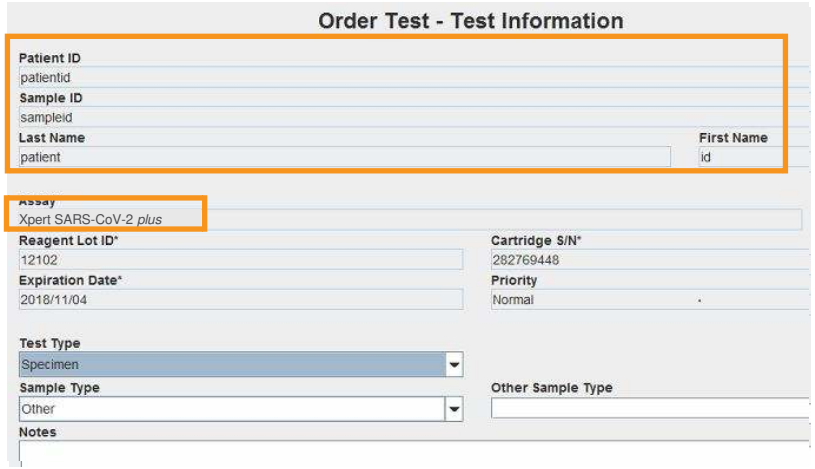
Run a Test on GeneXpert® Infinity (continued)

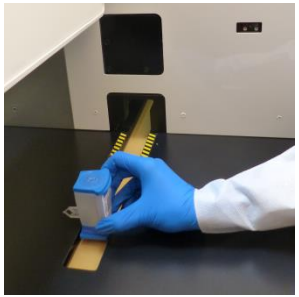
4 Complete the fields as required.

5 Select the assay protocol.

6 Click SUBMIT.

7 Place the cartridge onto the conveyor belt.





Automated Xpert® Xpress CoV-2/Flu/RSV *plus*

1. Sample is added to the cartridge
2. The cartridge is loaded into the System
3. Nucleic acids are purified
4. Purified nucleic acids mix with the PCR reagents
5. Simultaneous amplification and detection occurs
6. Results are ready to view

Cepheid.

Waste Disposal

- Biological specimens, transfer devices, and used cartridges should be considered capable of transmitting infectious agents and require use of standard precautions.
- Follow your institution's environmental waste procedures for proper disposal of used cartridges and unused reagents. These materials may exhibit characteristics of chemical hazardous waste requiring specific national or regional disposal procedures.
- If national or regional regulations do not provide clear direction on proper disposal, biological specimens and used cartridges should be disposed per WHO [World Health Organization] medical waste handling and disposal guidelines.

Quality Controls

Xpert® Xpress CoV-2/Flu/RSV *plus* Quality Controls

- Each Xpert cartridge is a self-contained test device.
- Cepheid designed specific molecular methods to include internal controls that enable the system to detect specific failure modes within each cartridge.
  - Probe Check Controls (PCC)
  - Sample Processing Control (SPC)

Refer to 301-4868 GeneXpert Quality Control features for all Cepheid assays.

Internal Quality Controls

- **Probe Check Controls (PCC)**
  - Before the PCR step, fluorescence signal is measured on all probes and compared with default factory settings to monitor:
    - Reagent rehydration
    - PCR tube filling
    - Probe integrity
    - Dye stability
- **Sample Processing Controls (SPC)**
  - SPC ensures that the sample was processed correctly and verifies that sample processing was adequate.
    - Verifies adequate extraction and amplification of the sample
    - Detects PCR inhibition
    - Ensures appropriate PCR conditions for amplification
    - Verifies that PCR reagents are functional
    - Must be positive in a negative sample to be a valid test
    - Can be positive or negative in a positive sample

Commercially Available External Controls

Zeptometrix	Description	Configuration	Storage
NATFRC-6C	Positive Control	6 x 0.5 mL	2-8°C or -20°C
NATCV9-6C	Negative Control	6 x 0.5 mL	2-8°C or -20°C

1. Open the cartridge lid.
2. Rapidly invert the external control tube 5 times.
3. Using a clean transfer pipette, transfer one draw (300 µL) of the external control sample into the large opening (Sample Chamber) in the cartridge.
4. Close cartridge lid.

- To minimize degradation of the control material, return any unused sample to the recommended storage conditions immediately after use.
- Many other vendors for quality control material are also available in addition to the one outlined above.
- External controls should be used in accordance with local, state accrediting organizations, as applicable.

Result Interpretation

Assay Targets

- SARS-CoV-2
- Flu A1
- Flu A2
- Flu B
- RSV
- SPC

Early Assay Termination

- The Xpress SARS-CoV-2 *plus* test mode includes an Early Assay Termination (EAT) function that will provide earlier time to result in high titer specimens if the signal from the SARS-CoV-2 target reaches a predetermined threshold before the full 45 PCR cycles have been completed.
- When SARS-CoV-2 titers are high enough to initiate the EAT function, the SPC amplification curve may not be seen and its results may not be reported.

Results Summary SARS-CoV-2 ADF

Result Displayed	SARS- CoV-2	SPC
SARS-CoV-2 POSITIVE	+	+/-
SARS-CoV-2 NEGATIVE	-	+
INVALID	-	-
ERROR	NO RESULT	NO RESULT
No Result	NO RESULT	NO RESULT

Results Summary SARS-CoV-2 and Flu ADF

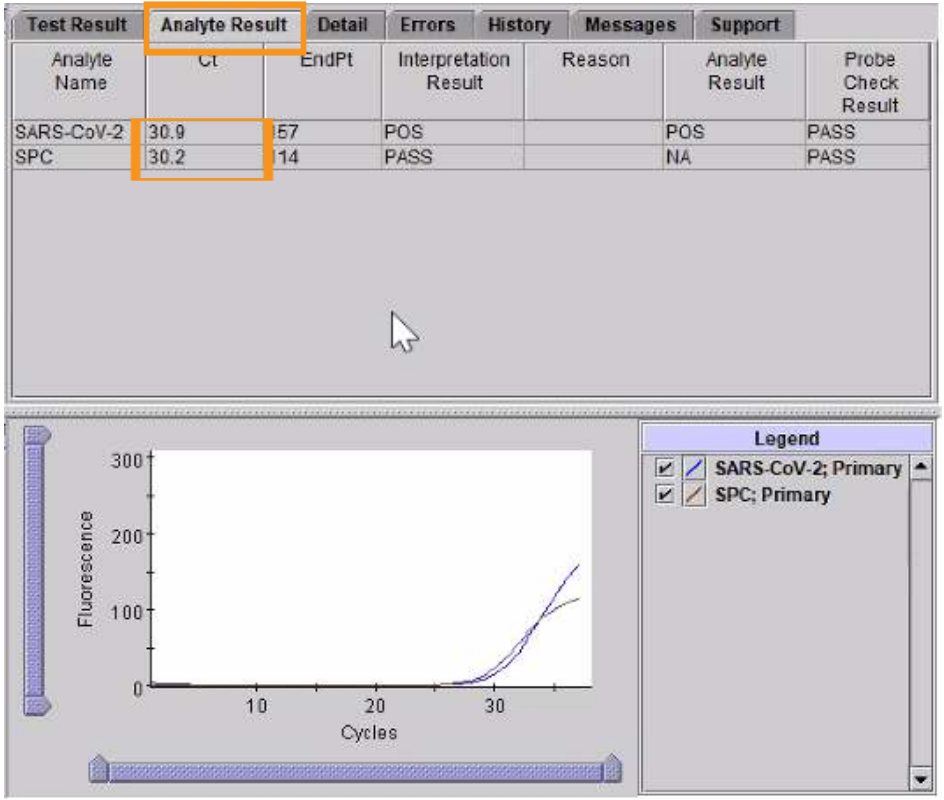
Result Displayed	SARS-CoV-2	Flu A1	Flu A2	Flu B	SPC
SARS-CoV-2 POSITIVE	+	-	-	-	+/-
Flu A POSITIVE	-	+	+/-	-	+/-
Flu A POSITIVE	-	+/-	+	-	+/-
Flu B POSITIVE	-	-	-	+	+/-
SARS-CoV-2 NEGATIVE Flu A NEGATIVE Flu B NEGATIVE	-	-	-	-	+
INVALID	-	-	-	-	-
ERROR	NO RESULT	NO RESULT	NO RESULT	NO RESULT	NO RESULT
No Result	NO RESULT	NO RESULT	NO RESULT	NO RESULT	NO RESULT

Results Summary SARS-CoV-2, Flu, and RSV ADF

Result Displayed	SARS-CoV-2	Flu A1	Flu A2	Flu B	RSV	SPC
SARS-CoV-2 POSITIVE	+	-	-	-	-	+/-
Flu A POSITIVE	-	+	+/-	-	-	+/-
Flu A POSITIVE	-	+/-	+	-	-	+/-
Flu B POSITIVE	-	-	-	+	-	+/-
RSV POSITIVE	-	-	-	-	+	+/-
SARS-CoV-2 NEGATIVE Flu A NEGATIVE Flu B NEGATIVE RSV NEGATIVE	-	-	-	-	-	+
INVALID	-	-	-	-		-
ERROR	NO RESULT	NO RESULT	NO RESULT	NO RESULT	NO RESULT	NO RESULT
No Result	NO RESULT	NO RESULT	NO RESULT	NO RESULT	NO RESULT	NO RESULT

Cepheid

SARS-CoV-2 POSITIVE



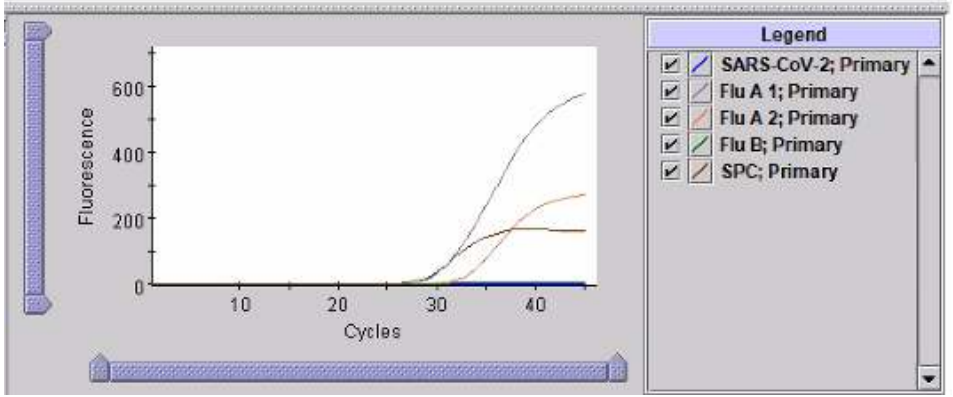
- SARS-CoV2- target RNA is detected
- SPC: NA; SPC is ignored because target amplification occurred
- Probe Check: PASS; all probe check results pass

SARS-CoV-2 Negative, Influenza A Positive, Influenza B Negative

Test Result	SARS-CoV-2 NEGATIVE; Flu A POSITIVE; Flu B NEGATIVE

Test Result | Analyte Result | Detail | Errors | History | Messages | Support

Analyte Name	Ct	EndPt	Interpretation Result	Reason	Analyte Result	Probe Check Result
SARS-CoV-2	0.0	2	NEG	No Ct	NEG	PASS
Flu A 1	30.3	576	POS		POS	PASS
Flu A 2	34.1	272	POS		POS	PASS
Flu B	0.0	7	NEG	No Ct	NEG	PASS
SPC	29.2	164	PASS		NA	PASS



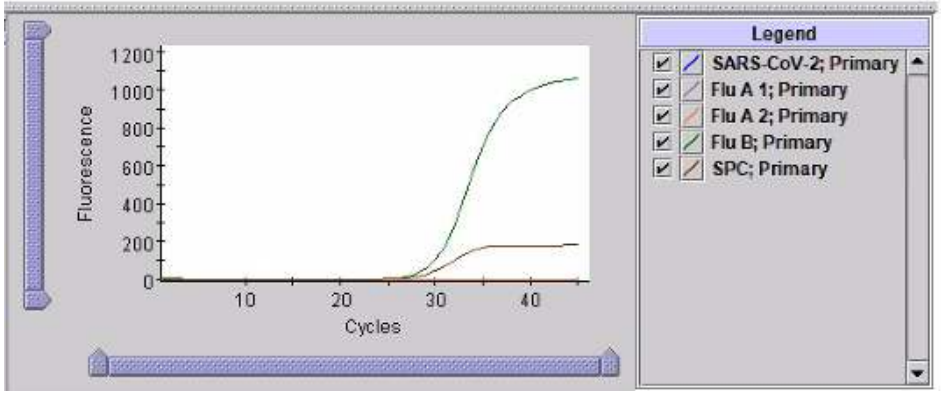
- SARS-CoV-2 not detected
  Flu A target RNA detected;
  Flu B target RNA not detected;
- SPC is ignored because target amplification occurred
- Probe Check: PASS; all probe check results pass

SARS-CoV-2 Negative, Influenza A Negative, Influenza B Positive

Test Result	SARS-CoV-2 NEGATIVE; Flu A NEGATIVE; Flu B POSITIVE

Test Result | Analyte Result | Detail | Errors | History | Messages | Support

Analyte Name	Ct	EndPt	Interpretation Result	Reason	Analyte Result	Probe Check Result
SARS-CoV-2	0.0	-6	NEG	No Ct	NEG	PASS
Flu A 1	0.0	-2	NEG	No Ct	NEG	PASS
Flu A 2	0.0	-1	NEG	No Ct	NEG	PASS
Flu B	28.5	1059	POS		POS	PASS
SPC	29.0	181	PASS		NA	PASS



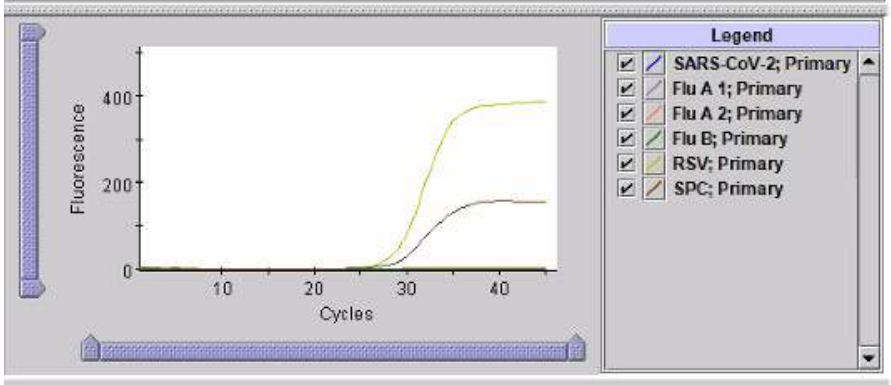
- SARS-CoV-2 not detected
  Flu A target RNA not detected;
  Flu B target RNA detected;
- SPC is ignored because target amplification occurred
- Probe Check: PASS; all probe check results pass

SARS CoV2 Negative, Influenza A Negative, Influenza B Negative, RSV Positive

Test Result	SARS-CoV-2 NEGATIVE; Flu A NEGATIVE; Flu B NEGATIVE; RSV POSITIVE

Test Result | Analyte Result | Detail | Errors | History | Messages | Support

Analyte Name	Ct	EndPt	Interpretation Result	Reason	Analyte Result	Probe Check Result
SARS-CoV-2	0.0	2	NEG	No Ct	NEG	PASS
Flu A 1	0.0	1	NEG	No Ct	NEG	PASS
Flu A 2	0.0	3	NEG	No Ct	NEG	PASS
Flu B	0.0	2	NEG	No Ct	NEG	PASS
RSV	29.0	389	POS		POS	PASS
SPC	29.6	157	PASS		NA	PASS

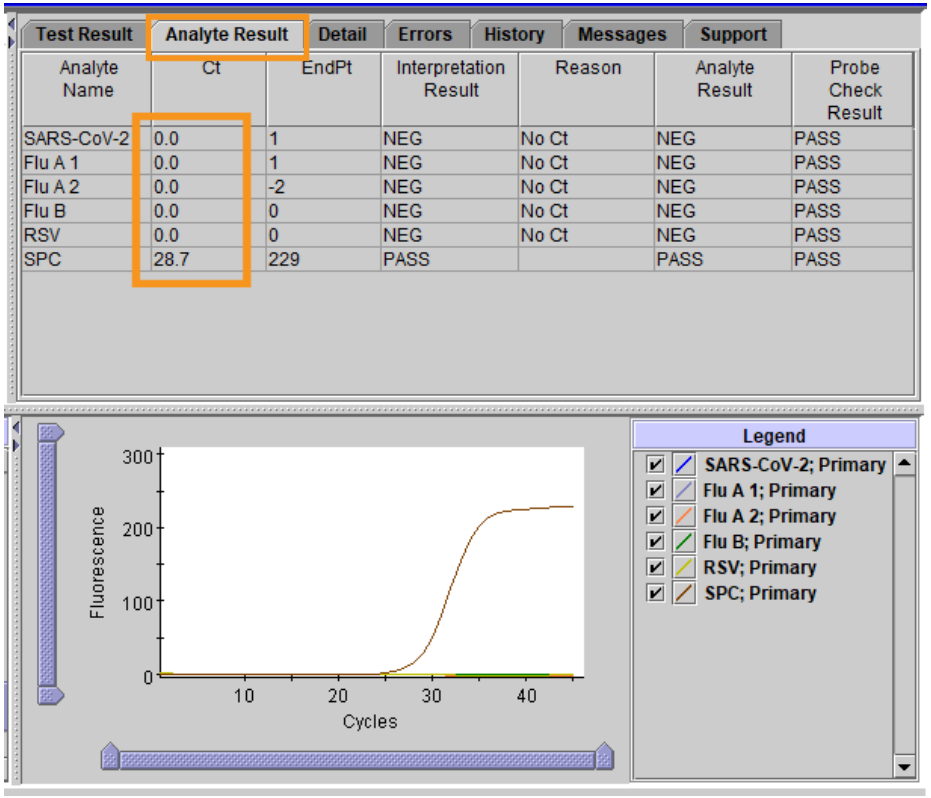


- SARS-CoV-2 not detected
  Flu A target RNA not detected
  Flu B target RNA not detected
  RSV target RNA detected
- SPC is ignored because target amplification occurred
- Probe Check: PASS; all probe check results pass

Cepheid

SARS CoV2 Negative, Influenza A Negative, Influenza B Negative, RSV Negative

Test Result	SARS-CoV-2 NEGATIVE; Flu A NEGATIVE; Flu B NEGATIVE; RSV NEGATIVE



- SARS-CoV-2 not detected
  Flu A target RNA not detected
  Flu B target RNA not detected
  RSV target RNA not detected
- SPC: PASS; SPC has a Ct within the valid range and endpoint above the minimum setting
- Probe Check: PASS; all probe check results pass

Limitations

- Performance of the Xpert® Xpress CoV-2/Flu/RSV *plus* test has only been established in nasopharyngeal and anterior nasal swab specimens. Use of the Xpert Xpress CoV-2/Flu/RSV *plus* test with other specimen types has not been assessed and performance characteristics are unknown.
- The performance of this test was established based on the evaluation of a limited number of clinical specimens. Clinical performance has not been established in all circulating variants but is anticipated to be reflective of the prevalent variants in circulation at the time and location of the clinical evaluation. Performance at the time of testing may vary depending on the variants circulating, including newly emerging strains of SARS-CoV-2 and their prevalence, which change over time.
- The performance of this device has not been assessed in a population vaccinated against COVID-19.

Limitations (continued)

- As with any molecular test, mutations within the target regions of the Xpert® Xpress CoV-2/Flu/RSV *plus* test could affect primer and/or probe binding resulting in failure to detect the presence of virus, or the virus being detected less predictably.
- This test cannot rule out diseases caused by other bacterial or viral pathogens.
- The performance of this test was validated using the procedures provided in this package insert only. Modifications to these procedures may alter the performance of the test.
- Erroneous test results might occur from improper specimen collection; failure to follow the recommended sample collection, handling, and storage procedures; technical error; or sample mix-up. Careful compliance with the instructions in this insert is necessary to avoid erroneous results.

Limitations (continued)

- False negative results may occur if virus is present at levels below the analytical limit of detection.
- Negative results do not preclude SARS-CoV-2, influenza or RSV infection and should not be used as the sole basis for treatment or other patient management decisions.
- Results from the Xpert® Xpress CoV-2/Flu/RSV *plus* test should be correlated with the clinical history, epidemiological data, and other data available to the clinician evaluating the patient.
- Viral nucleic acid may persist *in vivo*, independent of virus infectivity. Detection of analyte target(s) does not imply that the corresponding virus(es) are infectious or are the causative agents for clinical symptoms.
- This test has been evaluated for use with human specimen material only.
- This test is a qualitative test and does not provide the quantitative value of detected organism present.
- This test has not been evaluated for patients without signs and symptoms of respiratory tract infection.
- This test has not been evaluated for monitoring treatment of infection.

Limitations (continued)

- This test has not been evaluated for screening of blood or blood products for the presence of SARS-CoV-2, influenza, or RSV.
- The effect of interfering substances has only been evaluated for those listed within the labeling. Interference by substances other than those described can lead to erroneous results.
- Results from analytical studies with contrived co-infected samples showed potential for competitive interference of influenza B or RSV A at low concentrations (~3X LoD) when influenza A concentration is >1.7e5 RNA copies/mL or 1.7e6 RNA copies/mL, respectively. In addition, there is potential for competitive interference of influenza B at low concentration (~3X LoD) when SARS-CoV-2 concentration is >1e5 RNA copies/mL.
- Cross-reactivity with respiratory tract organisms other than those described herein can lead to erroneous results.
- Recent patient exposure to FluMist® or other live attenuated influenza vaccines may cause inaccurate positive results.

Limitations (continued)

- Zicam at 15% (w/v) may interfere with the detection of low levels of influenza B and RSV A.
- Samples collected into saline should not be frozen.
- As the Xpert® Xpress CoV-2/Flu/RSV *plus* test does not differentiate between the N2, RdRP and E gene targets, the presence of other coronaviruses in the B lineage, *Betacoronavirus* genus, including SARS-CoV-1 may cause a false positive result. None of these other coronaviruses is known to currently circulate in the human population.
- This test is not intended to differentiate RSV subgroups, influenza A subtypes or influenza B lineages. If differentiation of specific RSV or influenza subtypes and strains is needed, additional testing, in consultation with state or local public health departments, is required.
- Performance has not been established with media containing guanidine thiocyanate (GTC) other than eNAT.

Troubleshooting

Factors That Negatively Affect Results

- Improper specimen collection.
  - The performance of this assay with other specimen types or samples has not been evaluated.
- Inadequate numbers of organisms are present in the specimen.
- Improper transport or storage of collected specimen.
  - Storage and transport conditions are specimen specific.
  - Refer to the Instructions For Use for the appropriate handling instructions.
- Improper testing procedure.
  - Modification to the testing procedures may alter the performance of the test.
  - Careful compliance with the Instructions For Use is necessary to avoid erroneous results.

Reasons to Repeat the Test

- An **INVALID** result indicates that the control SPC failed. The sample was not properly processed, PCR is inhibited, or the sample was not properly collected.

- An **ERROR** result could be due to, but not limited to, Probe Check Control failure, system component failure, no sample added, or the maximum pressure limits were exceeded.

- A **NO RESULT** indicates that insufficient data were collected. For example, cartridge failed integrity test, the operator stopped a test that was in progress, or a power failure occurred.

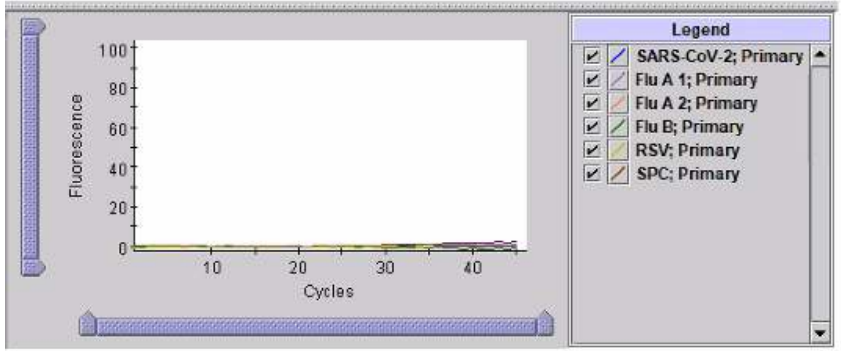
- If only one viral target is positive but coinfection with multiple targets is suspected, the sample should be re-tested with another FDA cleared, approved, or authorized test, if coinfection would change clinical management.

INVALID Result

Test Result	INVALID

Test Result | Analyte Result | Detail | Errors | History | Messages | Support

Analyte Name	Ct	EndPt	Interpretation Result	Reason	Analyte Result	Probe Check Result
SARS-CoV-2	0.0	1	NEG	No Ct	INVALID	PASS
Flu A 1	0.0	0	NEG	No Ct	INVALID	PASS
Flu A 2	0.0	1	NEG	No Ct	INVALID	PASS
Flu B	0.0	-1	NEG	No Ct	INVALID	PASS
RSV	0.0	0	NEG	No Ct	INVALID	PASS
SPC	0.0	3	FAIL	No Ct	FAIL	PASS



SPC does not meet acceptance criteria. Presence or absence of the target RNAs cannot be determined.

- SPC: FAIL;
- SARS-CoV-2, Flu A, Flu B, RSV signals do not have a Ct within valid range and endpoint below minimum setting
- Probe Check – PASS; all probe check results pass

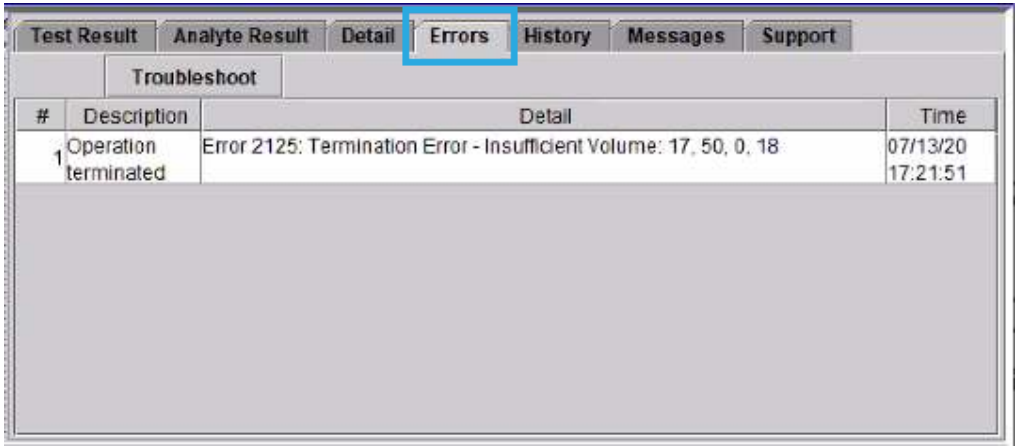
Possible Causes

- Improper sample collection or preparation
- Presence of interfering substances in the sample

Solution

- Repeat the test with a new cartridge

ERROR Result



Presence or absence of the target RNAs cannot be determined.

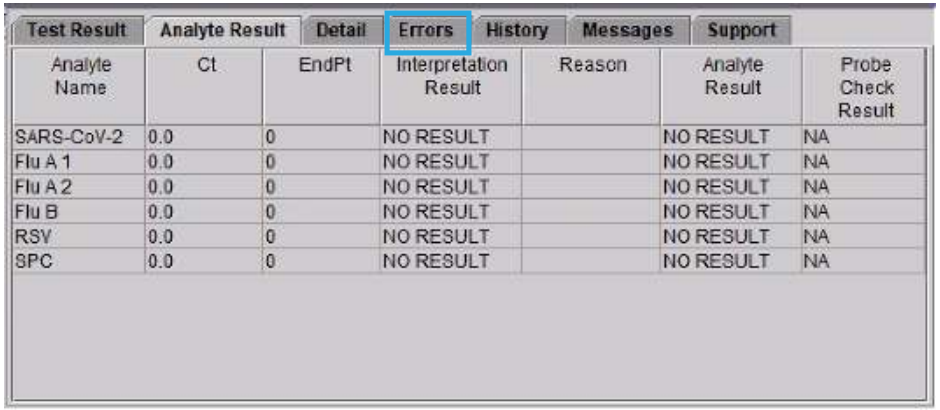
- SARS-CoV-2: NO RESULT
- Flu A: NO RESULT
- Flu B: NO RESULT
- RSV: NO RESULT
- SPC: NO RESULT
- Probe Check: FAIL; all or one of the probe check results fail

If the probe check passes, the error is caused by the maximum pressure limit exceeding the acceptable range or by a system component failure.

Solution

- Repeat the test with a new cartridge.

NO RESULT



Presence or absence of the target RNAs cannot be determined.

A **NO RESULT** indicates that insufficient data was collected. For example, the operator stopped a test that was in progress.

Possible Causes

A NO RESULT indicates that insufficient data was collected.

- Test was stopped with stop test button
- Electrical failure

Solution

- Secure the power
- Repeat the test with a new cartridge.

Retest Procedure

1

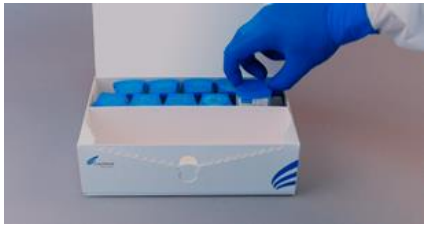
Discard used cartridge. Follow your institution's safety guidelines for disposal of cartridges.

2

Obtain the residual specimen. Prepare according to Instructions For Use.

If the leftover specimen volume is insufficient, or the retest returns an INVALID, ERROR, or NO RESULT, collect a new specimen.

3

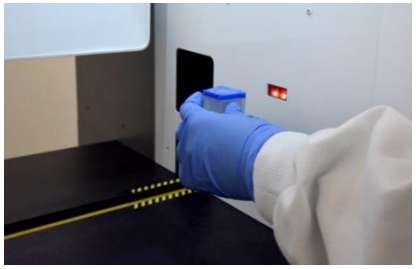


Obtain a new cartridge.

Process the specimen per the Instructions For Use.

4

Run the test on the system.



Technical Assistance

- Before contacting Cepheid Technical Support, collect the following information:
  - Product name
  - Lot number
  - Serial number of the System
  - Error messages (if any)
  - Software version and, if applicable, Computer Service Tag number

- Log your case online using the following link ***http://www.cepheid.com/en/support*** ***:*** *Create a Support Case*

Cepheid
A better way.

Thank You

www.cepheid.com